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(54) Title: ENHANCEMENT OF GENE EXPRESSION

(57) Abstract

A method for enhancing the expression of a selected gene in an organism while avoiding or reducing co-suppression involves the synthesis of a DNA which is altered in nucleotide sequence and is capable of expression of a protein, ideally identical to that of a protein already expressed by a DNA already present in the organism. This method ensures that sequence similarity between the two genes is reduced enough to eliminate the phenomenon of co-suppression, allowing the over-expression of a specific protein. The method is particularly suitable in plants.

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ENHANCEMENT OF GENE EXPRESSION

This invention relates to a method and material for enhancing gene expression in organisms, particularly in plants. One particular, but not exclusive, application of the invention is the enhancement of caroteniod biosynthesis in plants such as tomato (Lycopersicon spp.)

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In order to increase production of a protein by an organism, it is known practice to insert into the genome of the target organism one or more additional copies of the protein-encoding gene by genetic transformation. Such copies would normally be identical to a gene which is already present in the plant or, alternatively, they may be identical copies of a foreign gene. In theory, multiple gene copies should, on expression cause the organism to produce the selected protein in greater than normal amounts, this is referred to as "overexpression". Experiments have shown however, that low expression or no expression of certain genes can result when multiple copies of the gene are present. (Napoli et al 1990 and Dorlhac de Borne et al 1994). This phenomenon is referred to as co-suppression. It most frequently occurs when recombinant genes are introduced into a plant already containing a gene similar in nucleotide sequence. It has also been observed in endogenous plant genes and transposable elements. The effects of co-suppression are not always immediate and can be influenced by developmental and environmental factors in the primary transformants or in subsequent generations.

The general rule is to transform plants with a DNA sequence the codon usage of which approximates to the codon frequency used by the plant. Experimental analysis has shown that introducing a second copy of a gene identical in sequence to a gene already in the plant genome can result (in some instances) with the expression of the transgene, endogenous gene or both genes being inactivated (co-suppression). The mechanisms of exactly how co-suppression occurs are unclear, however there are several theories incorporating both pre- and post-gene transcriptional blocks.

As a rule the nucleotide sequence of an inserted gene is "optimised" in two respects. The codon usage of the inserted gene is modified to approximate to the preferred codon usage of the species into which the gene is to be inserted. Inserted genes may also be optimised in respect of the nucleotide usage with the aim of

approximating the purine to pyrimidine ratio to that commonly found in the target species. When genes of bacterial origin are transferred to plants, for example, it is well known that the nucleotide usage has to be altered to avoid highly adenylated regions, common in bacterial genes, which may be misread by the eukaryotic expression machinery as a polyadenylation signal specifying termination of translation, resulting in truncation of the polypeptide. This is all common practice and is entirely logical that an inserted sequence should mimic the codon and nucleotide usage of the target organism for optimum expression.

An object of the present invention is to provide means by which co-suppression may be obviated or mitigated.

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According to the present invention there is provided a method of enhancing expression of a selected protein by an organism having a gene which produces said protein, comprising inserting into a genome of the said organism a DNA the nucleotide sequence of which is such that the RNA produced on transcription is different from but the protein produced on translation is the same as that expressed by the gene already present in the genome.

The invention also provides a gene construct comprising in sequence a promoter which is operable in a target organism, a coding region encoding a protein and a termination signal characterised in that the nucleotide sequence of the said construct is such that the RNA produced on transcription is different from but the protein produced on translation is the same as that expressed by the gene already present in the genome.

The inserted sequence may have a constitutive promoter or a tissue or developmental preferential promoter.

It is preferred that the promoter used in the inserted construct be different from that used by the gene already present in the target genome. However, our evidence suggests that it may be sufficient that the region between the transcription and translation initiation codons, sometimes referred to as the "5" intervening region", be different. In other words, the co-suppression phenomenon is probably associated with the transcription step of expression rather than the translation step: it occurs at the DNA or RNA levels or both.

The invention further provides transgenic plants having enhanced ability to express a selected gene and seed and propagating material derived from the said plant.

This invention is of general applicability to the expression of genes but will be illustrated in one specific embodiment of our invention by a method of enhancing expression of the phytoene synthase gene which is necessary for the biosynthesis of carotenoids in plants, the said overexpression being achieved by the use of a modified transgene having a different nucleotide sequence from the endogenous sequence.

Preferably said modified phytoene synthase gene has the sequence SEQ-ID-10 NO-1.

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The invention also provides a modified chloroplast targeting sequence comprising nucleotides 1 to 417 of SEQ-ID-NO-1.

In simple terms, our invention requires that protein expression be enhanced by inserting a gene construct which is altered, with respect to the gene already present in the genome, by maximising the dissimilarity of nucleotide usage while maintaining identity of the encoded protein. In other words, the concept is to express the same protein from genes which have different nucleotide sequences within their coding region and, preferably the promoter region as well. It is desirable to approximate the nucleotide usage (the purine to pyrimidine ratio) of the inserted gene to that of the gene already present in the genome. We also believe it to be desirable to avoid the use of codons in the inserted gene which are uncommon in the target organism and to approximate the overall codon usage to the reported codon usage for the target genome.

The degree to which a sequence may be modified depends on the frequency of degenerate codons. In some instances a high proportion of changes may be made, particularly to the third nucleotide of a triplet, resulting in a low DNA (and consequently RNA) sequence homology between the inserted gene and the gene already present while in other cases, because of the presence of unique codons, the number of changes which are available may be low. The number of changes which are available can be determined readily by a study of the sequence of the gene which is already present in its degeneracy.

To obtain the gene for insertion in accordance with this invention it may be necessary to synthesise it. The general parameters within which the nucleotide sequence of the synthetic gene compared with the gene already present may be selected are:

- 5 1. Minimise the nucleotide sequence similarity between the synthetic gene and the gene already present in the plant genome;
 - 2. Maintain the identity of the protein encoded by the coding region;
 - Maintain approximately the optimum codon usage indicated for the target genome;
- 10 4. Maintain approximately the same ratio of purine to pyrimidine bases; and
 - 5. Change the promoter or, at least, the 5'-intervening region.

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We have worked with the phytoene synthase gene of tomato. The DNA sequence of the endogenous phytoene sequence is known (EMBL Accession Number Y00521): and it was discovered that this gene contained two sequencing errors toward the 3' end. These errors were corrected in the following way (1) cancel the cytosine at location 1365 and (2) insert a cytosine at 1421. The corrected phytoene synthase sequence (Bartley et al 1992), is given herein as SEQ-ID-NO-2. Beginning with that natural sequence we selected modifications according to the parameters quoted above and synthesised the modified gene which we designated MTOM5 and which has the sequence SEQ-ID-NO-1. Figure 1 herewith shows an alignment of the natural and synthesised gene with retained nucleotides indicated by dots and alterations by dashes. The modified gene MTOM5 has 63% homology at the DNA level, 100% at the protein level and the proportion of adenine plus thymidine (i.e. the purines) is 54% in the modified gene compared with 58% for the natural sequence.

In the sequence listings provided herewith, SEQ ID NO 1 is the DNA sequence of the synthetic (modified TOM5) gene rewferred to as MTOM5 in Figure 1, SEQ ID NO 2 is the natural genomic phytoene synthase (PsyI) gene referred to as GTOM5 in Figure 1, and SEQ-ID NO 3 is the translation product of both GTOM5 and MTOM5.

In tomato (Lycopersicon esculentum), it has been shown that the carotenoid namely lycopene, is primarily responsible for the red colouration of developing fruit (Bird et al 1991). The production of an enzyme phytoene synthase, referred to herein as PsyI, is an important catalyst in the production of phytoene, a precursor of lycopene.

Psyl catalyses the conversion of geranyl geranyl diphosphate to phytoene, the first dedicated step in carotenoid biosynthesis.

The regulation and expression of the active PsyI gene is necessary for the production of lycopene and consequently the red colouration of fruit during ripening. This can be illustrated by the yellow flesh phenotype of tomato fruits observed in a naturally occurring mutant in which the PsyI gene is inactive. In addition transgenic plants containing an antisense PsyI transgene, which specifically down regulates PsyI expression have also produced the yellow flesh phenotype of the ripe fruit.

When transgenic plants expressing another copy of the Psyl gene (referred to as TOM5) placed under the control of a constitutive promoter (being the Cauliflower Mosaic Virus 35S promoter) were produced, approximately 30% of the primary transformants produced mature yellow fruit indicative of the phenomenon of cosuppression. Although some of the primary transformants produced an increased caroteniod content, subsequent generations did not exhibit this phenotype thus providing evidence that co-suppression is not always immediate and can occur in future generations.

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The sequence of Psyl is known and hence the amino acid sequence was determined.

With reference to published Genbank genetic sequence data (Ken-nosuke Wada et al 1992.), a synthetic DNA was produced by altering the nucleotide sequence to one which still had a reasonable frequency of codon use in tomato, and which retained the amino acid sequence of Psyl. A simple swap between codons was used in cases where there are only two codon options, however in other cases the codons were changed within the codon usage bias of tomato. Nucleotide sequence analysis indicated that the synthetic DNA has a nucleotide similarity with Psyl (TOM5 Bartley et al 1992) of 63% and amino acid sequence similarity of 100%.

The synthetic gene was then cloned into plant transformation vectors under the control of 35S promoter. These were then transferred into tomato plants by Agrobacterium transformation, and both the endogenous and the synthetic gene appear to express the protein. Analysis of the primary transformants illustrates there is no evidence, such as the production of yellow fruit, indicative of co-suppression between the two genes.

The present invention will now be described by way of illustration in the following examples.

EXAMPLE 1

The coding region of the cDNA which encodes tomato phytoene synthase,

TOM5 (EMBL accession number Y00521) was modified since the original sequence
contained two errors towards the 3' end of the sequence. The sequence reported by
Bartley et al 1992 (J Biol Chem 267:5036-5039) for TOM5 cDNA homologues
therefore differs from TOM5 (EMBL accession number Y00521). For the purpose of
the production of the synthetic gene the sequence used is a corrected version of the
TOM5 cDNA which is identical to PsyI (Bartley et al 1992).

Design of the sequence.

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- Potential restriction endonuclease cleavage sites were considered given the
 constraints of the amino acid sequence. Useful sites around the predicted target
 sequence cleavage site were introduced to aid subsequent manipulation of the
 leader.
- 2. A simple swap between codons was used in cases where there are only two codon options (eg. lysine). In other cases codons were changed within the codon usage bias of tomato as given by Ken-nosuke Wada et al (codon usage tabulated from GenBank genetic sequence data, 1992. Nucleic Acids research 20:S2111-2118). A priority was given to reducing homology and avoiding uncommon codons rather than producing a representative spread of codon usage.
- 3. A BamHI site was introduced at either end of the sequence to facilitate cloning into the initial. At the 5' end 4A were placed upstream of the ATG according the dicot start site consensus sequence (Cavener and Ray 1991, Eukaryotic start and stop translation sites. NAR 19: 3185-3192).
- 4. The synthetic gene has been cloned into the vector pGEM4Z such that it can be translated using SP6.
- 5. Restriction site, stemloop and codon usage analyses were performed, all results30 being satisfactory.
 - 6. The modified TOM5 sequence was termed CGS48 or MTOM5.

Sequence analysis

CGS48

AT content = 54%

TOM5

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AT content = 58%

The nucleotide homology between TOM5 and CGS48 is 63%.

5 Amino acid sequence homology is 100%.

In summary the sequence TOM5 (Acc. No. Y00521) was extracted from the GenBank database and modified to incorporate the following corrections: deleted C at 1365, inserted C at 1421. CGS48 is based on the CDS of the modified Y00521 and the original sequence, whilst retaining translation product homology and trying to maintain optimal tomato codon usage.

Assembly of CGS48

CGS48 was divided into three parts:

CGS48A: BamHI / KpnI

CGS48B: KpnI / SacI

15 CGS48C: SacI / BamHI

All three were designed to be cloned on EcoRI / HindIII fragments. The sequences were divided into oligonucleotide fragments following computer analysis to give unique complementarity in the overlapping regions used for the gene assembly.

The oligonucleotides were synthesised on an Applied Biosystems 380B DNA synthesiser using standard cyanoethyl phosphoramidite chemistry. The oligonucleotides were gel purified and assembled into full length fragments using our own procedures.

The assembled fragments were cloned into pUC18 via their EcoRI/HindIII overhangs.

Clones were sequenced bi-directionally using "forward" and "reverse" sequencing primers together with the appropriate "build" primers for the top and bottom strands, using the dideoxy-mediated chain termination method for plasmid DNA.

Inserts from correct CGS48A, B and C clones were isolated by digestion with BamHI / KpnI, KpnI / SacI, SacI / BamHI respectively. The KpnI and SacI ends of the BamHI / KpnI and SacI / BamHI fragments were phosphatased. All three fragments were co-ligated into BamHI cut and phosphatased pGEM4Z. Clones with the correct sized inserts oriented with the 5' end of the insert adjacent to the SmaI site were

identified by PCR amplification of isolated colonies and digestion of purified plasmid DNA with a selection of restriction enzymes.

A CsCl purified plasmid DNA preparation was made from one of these clones. This clone (CGS48) was sequenced bi-directionally using "forward" and "reverse" sequencing primers together with the appropriate "build" primers for the top and bottom strands, using the dideoxy-mediated chain termination method for plasmid DNA.

EXAMPLE 2

Construction of the MTOM 5 vector with the CaMV 35S promoter

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The fragment MTOM5 (CGS48) DNA described in EXAMPLE 1 was cloned into the vector pJR1Ri (Figure 2) to give the clone pRD13 (Figure 3). The clone CGS48 was digested with Smal and Xbal and then cloned into pJR1Ri which was cut with Smal and Xbal to produce the clone pRD13.

EXAMPLE 3

Generation and analysis of plants transformed with the vector pRD13

The pRD13 vector was transferred to Agrobacterium tumefaciens LBA4404 (a micro-organism widely available to plant biotechnologists) and used to transform tomato plants. Transformation of tomato stem segments followed standard protocols (e.g. Bird et al Plant Molecular Biology 11, 651-662, 1988). Transformed plants were identified by their ability to grow on media containing the antibiotic kanamycin. Forty nine individual plants were regenerated and grown to maturity. None of these plants produced fruit which changed colour to yellow rather than red when ripening. The presence of the pRD13 construct in all of the plants was confirmed by polymerase chain reaction analysis. DNA blot analysis on all plants indicated that the insert copy number was between one and seven. Northern blot analysis on fruit from one plant indicated that the MTOM5 gene was expressed. Six transformed plants were selfed to produce progeny. None of the progeny plants produced fruit which changed colour to yellow rather than red during ripening.

The results are summarised in Table 1 below. The incidence of yellow, or mixed yellow/red (for example, striped) fruits is indicative of suppression of phytoene synthesis. Thus, with the normal GTOM5 construct, 28% of the transgenic plants displayed the co-suppressed phenotype. All the plants carrying the modified MTOM5

construct of this invention had red fruit demonstrating that no suppression of phytoene synthesis had occurred in any of them.

TABLE 1

	Construct			
	35S-GTOM5-nos	35S-MTOM5-nos		
Total number of fruiting plants	39	49		
Number of plants producing yellow fruit	8	0		
Number of plants producing mixed yellow and red fruit or temporal changes	3	0		
Number of plants producing red fruit	28	49		
% plants showing co-suppression of psyl	28%	0%		

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FIGURE 1

Sequence Alignment of Modified TOM5 with the synthetic MTOM5

10				W16	II the	3 y 11 t 11 t	CUIC IV	11011				
	TOM5	ATG	TCT	GTT						GTT	TCT	30
15	MTOM5	ATG M	AGC S	GTG V		CTT			GTG V	GTG V	AGC S	30
	TOM5	CCT	TGT	GAC	GTC	TCA	AAT	GGG	ACA	AGT	TTC	60
20	MTOM5	CCA P	TGC C	GAT D	GTG V	AGT S	AAC N	GGC G	ACT T	TCA S		60
20	TOM5	ATG	GAA	TCA	GTC	CGG			AAC	CGT	TTT	90
	MTOM5	ATG M	_	AGT S			GAA E	GGT	AAT N	AGA R	TTC F	90
25	TOM5	TTT	GAT							TTG	GTG	120
	MTOM5	TTC	GAC D	AGT	TCT S	CGT	CAC	CGT	AAC		GTT V	120
30		F	ם								•	
	TOM5	TCC	AAT	GAG		ATC	AAT	AGA		GGT		150
	MTOM5	AGT S	AAC N	-		ATA I		AGG	GGA	GGA	GGT	150
35	TOM5	AAG	CAA	ACT	AAT	AAT	GGA	CGG	AAA	TTT	TCT	180
	MTOM5	AAA	CAG	ACA	AAC	AAC	GGT	AGA	AAG	TTC	TCA	180

		K	Q	T	N	N	G	R	K	F	S	
	TOM5									CCA		210
5	MTOM5		AGA	TCA		ATC	CTT	GCA	ACA	CCT	AGC	210
	TOM5									CAG		240
10	MTOM5		GAG		acť		ACT	AGC	GAG	CAA Q	ATG	240
	TOM5									GCA		270
15	MTOM5	GTG	TAC	GAC	GTC	GTA	CTT	CGT	CAA	GCT A	GCA	270
	TOM5							-		ACC		300
20	MTOM5		GTT	AAA		CAG	TTA	CGT	AGT	ACT T		300
	TOM5									CCT		330
25	MTOM5	GAA	CTT	GAG	GTT	AAA	CCT	GAC	ATT	CCA P	ATA	330
	TOM5		GGG							GAA	GCA	360
30	MTOM5	CCT		AAC	CTT		CTT	CTT	TCT	GAG		360
	TOM5	TAT								GCA		390
35	MTOM5			AGA		GGA	GAG	GTT	TGC	GCA A		390
	TOM5											420
40	MTOM5	TAC		AAA	ACC		AAT	TTG			ATG	420
	TOM5	CTA	ATG									
45	MTOM5	TTG L			CCA		AGG		CGT	GCA A	ATA	450
	TOM5	TGG	GCA	ATA	TAT	GTA	TGG	TGC	AGA	AGA	ACA	480
50	MTOM5	TGG	GCT A	ATT	TAC	GTT	TGG	TGT	AGG	CGT R	ACI T	480
	TOM5											1 510
55	MTOM5		GAG	TTA	GTG	GAC	GGA	CCT	AAT	GCT A	AG	r 510
	TOM5	TAT	ATT	ACC	CCG	GCA	GCC	TTA	GAT	AGG	TG	3 540

	MTOM5	TAC Y	ATA I	ACA	CCC P	GCT	GCT	CTT	GAC D	AGA	TGG	540
5	TOM5		AAT									57 0
	MTOM5	GAG E	AAC	CGT R	TTG	GAG E	GAC	GTG V	TTT	AAC N	GGC G	570
10	TOM5		CCA									600
	MTOM5	AGA	CCT P	TTC	GAT	ATG	TTG	GAC	GGA G	GCA	CTT	600
15	TOM5	TCC	GAT	ACA								630
	MTOM5		GAC	ACT	GTG	AGC		TTC			GAC	630
20	TOM5		CAG									660
	MTOM5	ATC	CAA Q	CCT	TTT	CGG	GAC	ATG	ATC	GAG	GGC	660
25	TOM5		CGT	ATG								690
	MTOM5	ATG M	AGA	ATG	GAT	CTT	CGT	AAG		CGT	TAT	690
30	TOM5		AAC									720
	MTOM5	AAG K	AAT	TTT F	GAT	GAG	TTG	TAT Y	TTG	TAC	TGC C	720
35	TOM5		TAT									750
	MTOM5	TAC Y	TAC	GTG	GCA	GGA	ACC	GTG	GGC G	CTT	ATG M	750
40	TOM5	AGT	GTT	CCA					GCC			780
	MTOM5	TCA S	GTG V	CCT	ATC	ATG	GGA	ATT	GCA	CCA	GAG	780
45	TOM5		AAG									810
	MTOM5	AGT	AAA K	GCT	ACT	ACT	GAA	TCT	GTT	TAC	ACC	810
50	TOM5	GCT										840
	MTOM5	GCA	GCA A	CTA	GCA	TTA	GGT	ATA	GCT	AAC	CAG	840
55	TOM5	TTA	ACT	AAC	ATA	CTC	AGA	GAT	GTT	GGA	GAA	870
	MTOM5											

		L	T	N	I	L	R	D	v	G	E	
	TOM5											900
5	MTOM5	GAC	GCA A	CGT		GGT G	CGT	GTG V	TAT	CTC	CCA P	900
	TOM5								GGT	CTA	TCC	930
10	MTOM5	CAG		GAG		GCT	CAA		GGA		AGT S	930
	TOM5	GAT	GAA	GAT	ATA	TTT	GCT	GGA	AGG	GTG	ACC	960
15	MTOM5	GAC D	GAG	GAC D		TTC	GCA	GGT G	CGT	GTT	ACA	960
	TOM5										CAA	990
20	MTOM5	GAC	AAG	TGG	AGG	ATT	TTC	ATG	AAA	AAG	CAG	990
	TOM5	ATA	CAT	AGG	GCA	AGA	AAG	TTC	TTT	GAT	GAG	1020
25	MTOM5	ATT	CAC	CGT	GCT A	CGT	AAA	TTT F	TTC	GAC	GAA	1020
	TOM5	GCA	GAG	AAA	GGC	GTG	ACA	GAA	TTG	AGC	TCA	1050
30	MTOM5	GCT	GAA	AAG	GGA G	GTT	ACT T	GAG	CTT	TCT	AGT S	1050
	TOM5	GCT	AGT						GCA	TCT	TTG	1080
35	MTOM5	GCA A	TCA S		TTT F			TGG W	GCC A		CTT L	1080
	TOM5	GTC	TTG	TAC	CGC	AAA	ATA	CTA	GAT	GAG	ATT	1110
40	MTOM5	GTG	CTC L	TAT	AGA R	AAG	ATT	TTG	GAC D	GAA E	ATC I	1110
									TTC	ACA	AAG	1140
45	MTOM5	GAG	GCT	AAC	GAT D	TAT	AAT	AAT	TTT F		AAA K	1140
												1170
50	MTOM5	CGT	GCT	TAC	GTT V	TCT	AAG	AGC	AAA	AAA	CTT	1170
												1200
55	MTOM5	ATC	GCT	CTT	CCA P	ATC	GCT	TAC	GCT	AAG	AGC S	1200
	TOM5				CCT							1230

	MTOM5	TTG L	GTT V	CCA P	CCA P	ACT T	aag K	ACA T	GCT A	AGC S	TTG L	1230
_	TOM5	CAA	AGA	TAA								1239
5	мтом5	CAG Q	AGG R	TGA								1239
10			Same Diffe			se						
15			SEQU PEIN			: :	63% 100%		OLO OLO			

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40

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1239 base pairs

(B) TYPE: nucleic acid(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: SYNTHETIC DNA

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ATGAGCGTGG CACTTCTTTG GGTGGTGAGC CCATGCGATG TGAGTAACGG CACTTCATTT 60

15

ATGGAGAGTG TGAGAGAAGG TAATAGATTC TTCGACAGTT CTCGTCACCG TAACCTTGTT 120

AGTAACGAAC GTATAAACAG GGGAGGAGGT AAACAGACAA ACAACGGTAG AAAGTTCTCA
20 180

GTTAGATCAG CAATCCTTGC AACACCTAGC GGTGAGAGAA CTATGACTAG CGAGCAAATG 240

25 GTGTACGACG TCGTACTTCG TCAAGCTGCA CTAGTTAAAC GTCAGTTACG TAGTACTAAC 300

GAACTTGAGG TTAAACCTGA CATTCCAATA CCTGGAAACC TTGGACTTCT TTCTGAGGCT 360

30

TACGACAGAT GCGGAGAGGT TTGCGCAGAA TACGCTAAAA CCTTCAATTT GGGTACCATG

TTGATGACAC CAGAAAGGCG TCGTGCAATA TGGGCTATTT ACGTTTGGTG TAGGCGTACT
35 480

GACGAGTTAG TGGACGGACC TAATGCTAGT TACATAACAC CCGCTGCTCT TGACAGATGG 540

40 GAGAACCGTT TGGAGGACGT GTTTAACGGC AGACCTTTCG ATATGTTGGA CGGAGCACTT 600

	AGTGACACTG TGAGCAATTT CCCTGTGGAC ATCCAACCTT TTCGGGACAT GATCGAGGGG
5	ATGAGAATGG ATCTTCGTAA GTCTCGTTAT AAGAATTTTG ATGAGTTGTA TTTGTACTGC
	TACTACGTGG CAGGAACCGT GGGCCTTATG TCAGTGCCTA TCATGGGAAT TGCACCAGAG
10	AGTAAAGCTA CTACTGAATC TGTTTACACC GCAGCACTAG CATTAGGTAT AGCTAACCAG
15	CTTACAAATA TCTTGAGGGA CGTGGGTGAG GACGCACGTA GGGGTCGTGT GTATCTCCCA
15	CAGGACGAGC TCGCTCAAGC TGGATTGAGT GACGAGGACA TTTTCGCAGG TCGTGTTACA
20	GACAAGTGGA GGATTTTCAT GAAAAAGCAG ATTCACCGTG CTCGTAAATT TTTCGACGAA
	GCTGAAAAGG GAGTTACTGA GCTTTCTAGT GCATCAAGGT TTCCAGTTTG GGCCAGCCTT
25	GTGCTCTATA GAAAGATTTT GGACGAAATC GAGGCTAACG ATTATAATAA TTTTACTAAA 1140
30	CGTGCTTACG TTTCTAAGAG CAAAAAACTT ATCGCTCTTC CAATCGCTTA CGCTAAGAGC 1200
30	TTGGTTCCAC CAACTAAGAC AGCTAGCTTG CAGAGGTGA 1239
35	(2) INFORMATION FOR SEQ ID NO:2:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 1239 base pairs
	(B) TYPE: nucleic acid
40	(C) STRANDEDNESS: double
70	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: DNA (genomic)

	(iii) HYPOTHETICAL: NO
5	(iv) ANTI-SENSE: NO
	(vi) ORIGINAL SOURCE: (A) ORGANISM: LYOPERSICON ESCULENTUM (TOMATO)
10	(vii) IMMEDIATE SOURCE: (B) CLONE: GTOM5 - PHYTOENE SYNTHASE GENE
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
15	ATGTCTGTTG CCTTGTTATG GGTTGTTTCT CCTTGTGACG TCTCAAATGG GACAAGTTTC
20	ATGGAATCAG TCCGGGAGGG AAACCGTTTT TTTGATTCAT CGAGGCATAG GAATTTGGTG
20	TCCAATGAGA GAATCAATAG AGGTGGTGGA AAGCAAACTA ATAATGGACG GAAATTTTCT
25	GTACGGTCTG CTATTTTGGC TACTCCATCT GGAGAACGGA CGATGACATC GGAACAGATG
	GTCTATGATG TGGTTTTGAG GCAGGCAGCC TTGGTGAAGA GGCAACTGAG ATCTACCAAT
30	GAGTTAGAAG TGAAGCCGGA TATACCTATT CCGGGGAATT TGGGCTTGTT GAGTGAAGCA
35	TATGATAGGT GTGGTGAAGT ATGTGCAGAG TATGCAAAGA CGTTTAACTT AGGAACTATG
	CTAATGACTC CCGAGAGAAG AAGGGCTATC TGGGCAATAT ATGTATGGTG CAGAAGAACA
	GATGAACTTG TTGATGGCCC AAACGCATCA TATATTACCC CGGCAGCCTT AGATAGGTGG 540

GAAAATAGGC TAGAAGATGT TTTCAATGGG CGGCCATTTG ACATGCTCGA TGGTGCTTTG 600

TCCGATACAG TTTCTAACTT TCCAGTTGAT ATTCAGCCAT TCAGAGATAT GATTGAAGGA
5 660

ATGCGTATGG ACTTGAGAAA ATCGAGATAC AAAAACTTCG ACGAACTATA CCTTTATTGT 720

10 TATTATGTTG CTGGTACGGT TGGGTTGATG AGTGTTCCAA TTATGGGTAT CGCCCCTGAA 780

TCAAAGGCAA CAACAGAGA CGTATATAAT GCTGCTTTGG CTCTGGGGAT CGCAAATCAA 840

TTAACTAACA TACTCAGAGA TGTTGGAGAA GATGCCAGAA GAGGAAGAGT CTACTTGCCT

CAAGATGAAT TAGCACAGGC AGGTCTATCC GATGAAGATA TATTTGCTGG AAGGGTGACC 20 960

GATAAATGGA GAATCTTTAT GAAGAAACAA ATACATAGGG CAAGAAAGTT CTTTGATGAG

25 GCAGAGAAAG GCGTGACAGA ATTGAGCTCA GCTAGTAGAT TCCCTGTATG GGCATCTTTG 1080

GTCTTGTACC GCAAAATACT AGATGAGATT GAAGCCAATG ACTACAACAA CTTCACAAAG 1140

30

40

15

AGAGCATATG TGAGCAAATC AAAGAAGTTG ATTGCATTAC CTATTGCATA TGCAAAATCT 1200

CTTGTGCCTC CTACAAAAAC TGCCTCTCTT CAAAGATAA

35 1239

- (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 402 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: unknown

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

5 (iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: LYOPERSICONN ESCULENTUM (TOMATO)

10 (vii) IMMEDIATE SOURCE:

(A) LIBRARY: TRANSLATION PRODUCT OF GTOM5 AND MTOM5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

15

Met Ser Val Ala Leu Leu Trp Val Val Ser Pro Cys Asp Val Ser Asn
1 5 10 15

Gly Thr Ser Phe Met Glu Ser Val Arg Glu Gly Asn Arg Phe Phe Asp 20 25 30

Ser Ser Arg His Arg Asn Leu Val Ser Asn Glu Arg Ile Asn Arg Gly 35 40 45

25 Gly Gly Lys Gln Thr Asn Asn Gly Arg Lys Phe Ser Val Arg Ser Ala 50 55 60

Ile Leu Ala Thr Pro Ser Gly Glu Arg Thr Met Thr Ser Glu Gln Met 65 70 75 80

30

Val Tyr Asp Val Val Leu Arg Gln Ala Ala Leu Val Lys Arg Gln Leu 85 90 95

Arg Ser Thr Asn Glu Leu Glu Val Lys Pro Asp Ile Pro Ile Pro Gly
100 105 110

Asn Leu Gly Leu Leu Ser Glu Ala Tyr Asp Arg Cys Gly Glu Val Cys 115 120 125

40 Ala Glu Tyr Ala Lys Thr Phe Asn Leu Gly Thr Met Leu Met Thr Pro
130 135 140

SUBSTITUTE SHEET (RULE 26)

Glu Leu Ser Ser Ala Ser Arg Phe Pro Val Trp Ala Ser Leu Val Leu

Tyr Arg Lys Ile Leu Asp Glu Ile Glu Ala Asn Asp Tyr Asn Asn Phe

Thr Lys Arg Ala Tyr Val Ser Lys Ser Lys Lys Leu Ile Ala Leu Pro 370 375 380

Ile Ala Tyr Ala Lys Ser Leu Val Pro Pro Thr Lys Thr Ala Ser Leu

5 385 390 395 400

Gln Arg

CLAIMS

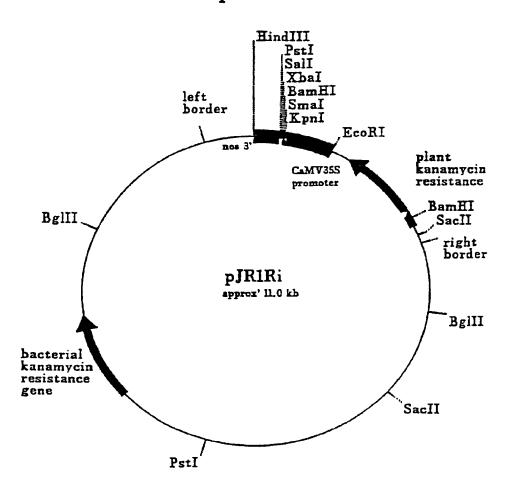
- A method of enhancing expression of a selected protein by an organism having a gene which produces said protein, comprising inserting into the genome of the said organism a DNA the nucleotide sequence of which is such that the RNA produced on transcription is different from but the protein produced on translation is the same as that expressed by the gene already present in the genome.
- 10 2. A method as claimed in claim 1, in which the organism is a plant.
 - 3. A method as claimed in claim 2, in which the plant is a tomato plant.
- 4. A method as claimed in any preceding claim, in which the selected gene is the gene encoding phytoene synthase.
 - 5. A method as claimed in claim 4, in which the coding region of the said inserted gene has the sequence SEQ-ID-NO-1.
- 20 6. A gene construct comprising in sequence a promoter which is operable in a target organism, a coding region encoding a protein and a termination signal characterised in that the nucleotide sequence of the said construct is such that the RNA produced on transcription is different from but the protein produced on translation is the same as that expressed by the gene already present in the genome.
 - 7. A method of enhancing expression of caroteniods in a plant comprising overexpression in the plant a gene specifying an enzyme necessary to the biosynthesis of carotenoids, the said overexpression being achieved by the use of a modified transgene having a different nucleotide sequence from the endogenous sequence.

- 8. A method as claimed in claim 7, in which the modified gene specifies phytoene synthase.
- 9. A modified chloroplast targeting sequence comprising nucleotides 1 to 417 of
 5 SEQ-ID-NO-1

1/2

FIGURE 2

pJR1Ri

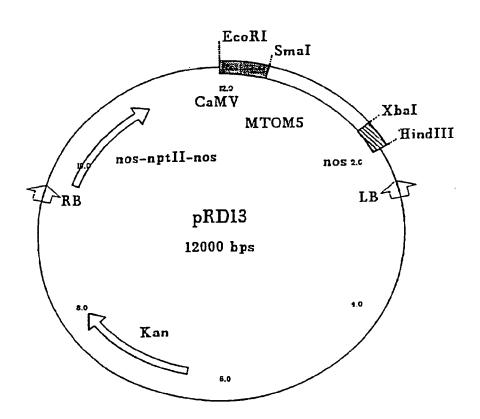


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2/2
FIGURE 3

pRD13

MTOM5 encodes phytoene synthase



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A. CLASSIF IPC 6	C12N15/67 C12N15/82 C12N15/29	C07K14/415	
According to	International Patent Classification (IPC) or to both national classification	and IPC	
B. FIELDS		vmhnis)	
Minimum do IPC 6	cumentation searched (classification system followed by classification of C12N C07K	, mose,	
Documentat	ion searched other than minimum documentation to the extent that such	documents are included in the fields search	hed
Electronic d	ale base consulted during the international search (name of date base	and, where practical, search terms used)	
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
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 Y	pages 693-701, XP002043131 see the whole document		1-4
Υ	WO 90 02189 A (UPJOHN CO) 8 March see the whole document	1990	1-4
X	WO 95 02060 A (ZENECA LTD ;GRIERS (GB); FRAY RUPERT GEORGE (GB)) 19 1995 see page 18, paragraph 1	ON DONALD January	1-4
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	10 October 1997	1 7. 10. 97	
Name an	d mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Maddox, A	

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	Relevant to claim No.
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FINNEGAN, J., ET AL.: "Transgene inactivation: plants fight back" BIOTECHNOLOGY, vol. 12, September 1994, pages 883-888, XP002043134 see the whole document	1-8
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